

Waringer and Humpesch, 1984; Corbet, 1999).

The purpose of this study was to follow the life cycle stages of *I. evansi* experimentally at different temperatures in laboratory. The results then, would be useful to applied for filed study.

MATERIALS AND METHOD:

The laboratory studies of the different life cycle aspects were based on fertilized females, captured from the field at a tandem condition. In laboratory, each female was inculated in a glass jar (500ml) supplied with a wited filter paper with water and stems of *Bacopa monniera* (L.) (aquatic plant) for a perch (Cordero, 1990). The mouth of the jars were closed with gauze and the females were watched until oviposition occurred.

The eggs describtion, dimention and development were done using dissecting microscope. When a female lay eggs on the filter paper, the eggs were divided into 10 replicates each with 30 eggs before placed in a plastic petri-dishes (5x 1.5 cm) containing a small quantity of water. Eggs development were investigated at several constant

temperatures (10, 12, 15, 20, 25, 30, 35, 40 and 43°C) in cooled incubator.

For larval stage development, several experiments were performed in cooled incubator by using 50 newly-hatched larvae (8h.old) for each experiment. Larvae were placed individually in 40 ml plastic tubes that were filled with tap water and a stem of the plant *Bacopa moniera* for a perch. The larvae were fed on *Artemia salina* nauplii (Cordero, 1990), when the larvae reach the instar five they were transferred to bigger tubes filled with 75ml water. The growth of successive instars were followd by detecting the larval moults and recording the duration time for each instar. The identification of different instars number were done according to Benke(1970), were the final instar=0, penultimate =1, antepenultimate =2, etc.

The morphometric measurements were done by using a micrometer fitted with the eye-piece of a dissecting microscope (reading were made to the nearest 0.01mm.). The head width was taken across the widest part of the head, usually from the tip of one eye to the tip of the other and the total body length was measured from the anteriormost point

of the head to the tip of the abdomen without caudal gills (Britt, 1953). The growth and larval development experiments were done at 3 constant temperatures (25, 35 and 40 ± 1 °C). Adults that emerge from the final larval instar were maintained in an aquarium supplied with *Drosophila* as food and they were observed until the appearance of the males and female colors (Cordero, 1990).

RESULTS:

Incubation period:

The effects of temperature on eggs hatching and incubation period are shown in (Table 1). The temperatures values 12 and 40 °C

were found to be the lower and upper limits of eggs hatching, respectively. However the percentages of hatching were positively increased with increasing temperature and the optimum hatching was recorded at 35°C.

The incubation period was inversely related to temperature. The period was reduced from 31 days at 12 °C to only 7 days at 40 °C and the ANOVA test showed that the differences is highly significant ($F_{(6,28)} = 41.9, P < 0.001$). Furthermore the differences were significant between 12 and 25°C and between 25 and 40°C ($P < 0.05$).

Table (1): *I. evansi*, laboratory study. Incubation period and egg hatching at different temperatures (different letter represent significant differences, one – way ANOVA).

Temp. (°C)	Incubation period (days)	Hatching (%)
	(mean and range)	
10	- -	No hatching
12	a 31 (26-37)	40
15	a 26 (23-30)	67
20	ab 23 (20-27)	81
25	ab 20 (17-23)	96
30	b 14 (11-19)	99
35	b 10 (9-12)	100
40	c 7 (5-10)	43
43	- -	No hatching

The egg stage:

The egg of *I. evansi* is of cylindrical shape, white-yellowish become orange - brown after 48h. They are narrow and smooth at the front end having a funnel-shape, which is somewhat free. Its length was ranged from 0.71-0.78, $X=0.75$ mm and the diameter ranged from 0.18-0.22, $X=0.20$ mm (n= 52).

The larval development:

The first larval instar was moulted 12 times before emerged (Table 2) and therefore the insect completed through 13 instars. Moreover , the number of instars was remained constant at all tested temperatures (25,35 and 40 °C)

Instars duration times and temperature:

In general it appear that temperature had an effective role on the duration time of various instars. The mean duration time of instars (in days) were ranged from 1.0 ± 0.17 at 25 °C to 0.5 ± 0.01 at 40°C for instar (12) and from 69.9 ± 0.12 at 25°C to 58 ± 0.110 at 40°C for instar (0) (Table 2). It's obvious that temperature and the instar duration times are inversely correlated.

Instars duration times and growth

The duration time become longer as the larvae moulted and growing under all tested temperatures (Table 2). For example in instar (12) it takes 0.5-1.0 day at 40 and 25°C respectively . Whilest in instar (0) it takes 58.0-69.9 days at 40 and 25°C respectively. However, the general relationships between instars duration times and instars number (growth) can be represented by a linear correlation equations as given in (Table 3). The relations equations for the 3 examined temperatures (25,35 and 40°C) indicated that the duration time was increased directly as the larvae become older at all temperatures and the correlation constants (a and b) was increasing too.

Head size increments:

The head size increments represented by the increments of the head width of the larvae at different temperatures are shown in (Table.4) . In general the results showed that increments increased gradually with highest values of the last two instar (0.327-0.377mm).however ,despite that larvae reared at higher temperature, could have wider heads, for example the mean H.W of the

larvae at instar (12) was 0.37 mm. at 25 °C, 0.39 mm. at 35 °C and 0.40 mm at 40°C. For the final larval stage, the instar (0). The mean H.W.

were 2.89, 2.95 and 3.10 mm at 25, 35 and 40°C respectively, the statistical ANOVA test showed that differences is not significant.

Table (2): *I. evansi* (Garmat Ali, Basrah), laboratory study. Instar numbers and duration time at different temperatures. final instar = 0, penultimate = 1, antepenultimate = 2, etc.

Instar	Duration time (days)		
	25 °C	35 °C	40 °C
	(mean ± SD and range)	(mean ± SD and range)	(mean ± SD and range)
12	1.0 ± 0.171 (0.99-2.0)	1.0 ± 0.029 (0.99-2.0)	0.5 ± 0.011 (0.5-1.0)
11	7.6 ± 0.095 (7-16)	4 ± 0.090 (3-6)	2.8 ± 0.120 (2-4)
10	8 ± 0.103 (7-15)	4.4 ± 0.112 (3-6)	3.1 ± 0.70 (2-4)
9	8.3 ± 0.071 (6-21)	5.7 ± 0.192 (4-13)	4.25 ± 0.91 (3-7)
8	9.5 ± 0.05-75 (6-15)	7.3 ± 0.56 (5-12)	4.7 ± 0.022 (5-9)
7	10.7 ± 0.104 (9-12)	7.6 ± 0.171 (6-14)	5.2 ± 0.110 (3-8)
6	13 ± 0.100 (8-16)	9.4 ± 0.121 (6-18)	7.1 ± 0.142 (5-17)
5	15.7 ± 0.920 (14-17)	13.6 ± 0.056 (11-21)	8.5 ± 0.082 (5-13)
4	20.5 ± 0.090 (14-27)	18.8 ± 0.092 (14-28)	11.5 ± 0.012 (10-17)
3	30.3 ± 0.64 (24-24)	24 ± 0.187 (20-46)	22.5 ± 0.101 (31-17)
2	36 ± 0.48 (32-51)	31 ± 0.54 (27-45)	27 ± 0.099 (13-28)
1	48.2 ± 0.96 (39-57)	42.7 ± 0.090 (34-50)	36.5 ± 0.107 (29-41)
0	69.9 ± 0.124 (41-70)	62 ± 0.104 (44-73)	58 ± 0.110 (43-73)

Table (3): *I. evansi* (Garmat Ali, Basrah), laboratory study. Linear correlation equations between duration times and instars at 3 temperatures, $Y = a + bX$, Y is instars and X is duration time, a and b are constants.

Temp.(°C)	Relation	R	N	P
25	$Y = 3.23 + 0.17 X$	0.89	13	< 0.05
35	$Y = 3.66 + 0.19 X$	0.88	13	< 0.05
40	$Y = 4.03 + 0.197 X$	0.86	13	< 0.05

Table (4): *I. evansi* (Garmat Ali , Basrah) , laboratory study. Mean head width (mm) at different temperatures. No, number at speimaes final instar =0 penultimate =1 , antepenultimate =2 , etc. ,mean moult increment (Mo.)

Instars	Head width (mm)			Mo.
	25 °C	35 °C	40 °C	
	(mean ± SD and range and No)	(mean ± SD and range and No)	(mean ± SD and range and No)	
12	0.25 ± 0.016 (0.03-0.30) 50	0.25 ± 0.017 (0.23-0.30) 50	0.25 ± 0.020 (0.23-0.30) 50	
11	0.37 ± 0.025 (0.32 – 0.40) 50	0.39 ± 0.019 (0.30 – 0.42) 41	0.40 ± 0.021 (0.30 – 0.49) 50	0.137
10	0.47 ± 0.050 (0.40 – 0.55) 50	0.51 ± 0.05 (0.40 – 0.57) 39	0.54 ± 0.22 (0.47 – 0.69) 50	0.120
9	0.61 ± 0.065 (0.50 – 0.65) 50	0.64 ± 0.073 (0.55 – 0.80) 39	0.78 ± 0.012 (0.67-0.97)50	0.170
8	0.73 ± 0.069 (0.63 – 0.85) 48	0.86 ± 0.107 (0.63 – 1) 34	0.93 ± 0.11 (0.82 – 1.17) 50	0.163
7	0.85 ± 0.079 (0.75 – 0.97) 45	1.08 ± 0.065 (0.97 – 1.25) 34	1.22 ± 0.120 (1.10 – 1.45) 50	0.21
6	1.01 ± 0.094 (0.89 – 1.19) 45	1.26 ± 0.104 (1.12 – 1.50) 30	1.45 ± 0.110 (1.35 – 1.72) 42	0.19
5	1.24 ± 0.104 (1.12 – 1.42) 42	1.43 ± 0.112 (1.30 – 1.69) 30	1.70 ± 0.120 (1.52 – 1.89) 40	0.217
4	1.44 ± 0.108 (1.29 – 1.62) 42	1.64 ± 0.121 (1.37 – 1.85) 30	1.97 ± 0.164 (1.77 – 2.25) 36	0.227
3	1.64 ± 0.065 (1.60 – 1.77) 40	1.98 ± 0.214 (1.62 – 2.32) 29	2.29 ± 0.141 (2.17 – 2.52) 36	0.253
2	1.92 ± 0.012 (1.75 – 2.25) 39	2.27 ± 0.191 (2.10 – 2.72)25	2.62 ± 1.90 (2.54 – 2.90) 34	0.286
1	2.30 ± 0.192 (2.15 – 2.55) 37	2.62 ± 0.019 (2.44 – 2.87) 23	2.89 ± 0.097 (2.57 – 3.01) 34	0.327
0	2.89 ± 0.120 (2.25 – 3.01) 37	2.95 ± 0.131(2.75 – 3.12) 20	3.10 ± 0.99 (2.75 – 3.22) 34	0.377

Adults:

Laboratory observations showed that the adult insect emerged from the final larval instar by a departure on the dorsal plate of the head and thorax. The male of the insect were found to have one colour, they are only blue, whereas the females were polymorphous in colours and may be like male (homeochromes, andromorphs), brown (heterochromes, gynochrome) or orange-brown (heterochromes, androchromotomics).

DISCUSSION:

Under controlled conditions *I. evansi*, hatched at relatively wide range of temperatures (12-40°C) and showed a rapid embryonic development, particularly at the upper temperature limits. Hatching was completed in 1 day at 40°C. Generally lower embryonic growth, were reported for other species at different other regions. Waringer and Humpesch (1984) had reported that hatching of *Coenagrion puella* from lower Austria occurs within the temperature range 12-28°C. Furthermore, they mentioned to the results of other studies (Boehms, 1971; Sawchyn and Gillott, 1974;

Rivard *et al.*, 1975) for a number of species which have similar ranges (12-28°C) or slightly wider but however, their upper limits were not more than 30°C. Vertically this is 10°C less than. The shortest periods for embryonic development recorded in these studies were, 15 days in *Coenagrion resolutum*, 16 days in *C. angulatum*, 12 days in *C. puella*. Nevertheless, temperature has been shown to be the major factor determining the eggs development time in moulted environments (Corbet, 1960; Waringer and Humpesch, 1984). Obviously, the present results reflect the subtropical habitat of *I. evansi*.

After hatching, odonata could have a number of moults usually, ranged between seven to thirteen moults before the adult stages attained (Corbet, 1980). For *I. evansi* thirteen moults has been found to complete the larval development and this number was found to be constant at all studied temperature conditions (25, 35 and 40°C).

Actually the effects of temperature on growth may appear in different ways. In one way temperature has a direct positive effect on the size, so the larvae could

have larger size when hatched and reared at higher temperature. This relationship has been reported in many other species of damselfly such as, *I. elegans*, *C. resolutum*, *C. puella*, *Pyrrhosoma nymphula* (Lawton, 1971; Thompson, 1978; Baker and Clifford, 1981) and dragonfly such as *Aeschna mixa* and *Papopleura lucia* (Schaller, 1972; Hassan, 1976). From other way the duration time of the larval instars become shorter at rising the temperature as for example in this study when temperature rised from 25°C to 35°C and to 40°C. Actually the reverse relationship between temperature and intermoult period or the instar duration time is well documented for insects and crustacea, in general (Grodzinski, *et al.*, 1975; Downing and Rigler, 1984).

On the basis of the results of the present study the life cycle of *I. evensi* can be completed, under laboratory conditions, within 197 days at the upper limit of temperature (40°C) and within 278days at 25°C. Despite the period of development is less than one year it can not be concluded that the population of *I. evensi* is a semivoltine, before further investigation on the ecology of the insect is done in the field. But because of some European

Coenagrionidae had been found to have a semivoltine populations, e.g. *C. hastulatum* (Haritonov, 1977) and *C. resolutum* (Baker and Clifford, 1981) as well as many crustacean species, living in the same region *I. evansi* i.e in Shatt Al-Arab river system, have been found semivoltine (Ali and Salman, 1987; Salman, *et al.*, 1996), therefor it can be expected that the population of *I. evansi* is able to produce more than one generation during the year.

REFERENCES:

- Abdul-Karim, R.M.1994. New records of some dragonflies and damselflies (Odanata) from Basrah City, Iraq. Marina Mesoptamica. (1): 79-89.
- Al-Ali, A.S. 1977. Phytophagous and entomophagous insects and mites in Iraq. Publ.Nat. Hist. Res. Center. 33:142.
- Ali, M.H and Salman, S.D.1987. Growth and the amphipod *Parhyale basreneis* (Talitridae) in the Shatt Al-Arab region. Mar.Ecol. prog. Ser. 40:231-238.
- Ali, M.H; Annon, M.R. and Mohammed H.H. 2002.The seasonal variations of abundance and biomass of two

- odonates naiads *Ishnura evansi* morton (*Odonata: coenagrionidae*) and *Brachythemis fuscopalliata* selys (*Odonata: libellulidae*) at Garmat Ali region, Basrah .Marina Mesopotamica,17(2):405 – 415.
- Baker, R.L and Clifford, H.F.1981. Life cycles and food of *Coenagraion resolutum* (*Coenagrionidae : Odonata*) Population from the Boreal forset of Alberta. Can. Aguat. Insec. 3:179-191.
- Benke, A.C.1970 .A method for comparing individual growth rate of aquatic insects with special reference to the odonata. Ecology, 51: 328-331.
- Benke, A.C.1976. Dragonfly production and prey turnover. Ecology,57, 5:915-927.
- Boehms, Ch. N.1971. The influence of temperature upon embryonic diapause and seasonal regulation in *Sympetrum vicinum* (*Odonata: Libellulidae*). Entamology Gaz. 13:2-20.
- Britt, N.W.1953. Difference between measurement of living and preserved aquatic nymphs caused by injury and preservative. Ecology, 34: 802-804.
- Corbet, P.S., 1960. Pattrens of circadian rhythm in insects. Coldspring Harbor symp. Quant. Biol., New York 25:357-360.
- Corbet, P.S., 1980. Biology of dragonflies. Ann.Rev.Entomol.25: 189-217.
- Corbet, P.S., 1999.A biology of dragonfiles .3et. Cornell University Press. England
- Cordero, A.1990. The inheirtance of female polymorphism in the damselflies *Ischnura graellsii* (*Odonata:Coenagrionidae*). Heredity 64:341-346.
- Dowining, J.A. and Rigler, F.H.1984. A manual on methods for the assessment of secondary production in fresh water. IBP Hand Book No.17 Black well, Oxford.1-18.
- Grodzinski, W. Klekowski, R. and Duncan, A.1975. Methods for ecological bioenergetics. IBP Handbook No. 24. Blackwell, Oxford
- Haritonov, A. Y. 1977. Life cycles of some species of dragonflies in eastern priuralje. Ivestia Sibirskogo Odeleina Akademii Nauk. USSR, 5, 55 - 60
- Hassan, A.T. 1976.The effect of food on the larval development of

- Palpoleura lucia* (Anisoptera : Libellulidae). Odonatologica. 5:27-33.
- Khalaf,A.N. and Al-Omer,M.A.1974. A second list of insects from Iraq. Bio. Res. Center. 2:1-41.
- Lawton, J.H.1971. Maximum and actual feeding rates in larvae of the damselfly *Pyrhosoma nymphula* (Odonata: Zygoptera). Freshwater Biology 1:99-111.
- Rivard, D,Pilon, J.G. and Thiphakesone, S.1975. Effect of constant temperature environments on egg development of *Enallagma boreale* (Zygoptera: Coenagrionidae). Odonatologica., 4:271-276.
- Salman,D.S.Oshana, K.V. and Ali,M.H. 1996. Life cycle and population dynamics of *Annina mesopotamica* (Ahmed), (Isopoda: Flabellifera) in the Shatt Al-Arab region, Basrah, Iraq. Hydrobiologia 330:119-130.
- Sawchyn, W.W.and Gillott, C. 1974. The life history of *Lestes cogenes* on the Canadian prairies. Can. Entomol. 106:76-367.
- Schaller,F. 1972. Action for temperature and diapause embryonic development of *Aeshna mixia* (Aeshnidae: Odonata). Odonata., 1:143-153.
- Thompson, D.J.1978. Prey size selection by larvae of damselfly, *Ischnura elegans*. J.Anim. Ecol.,47:85-769.
- Waringer, J.A. and Humpesch, U.K.1984. Embryonic development, larval growth and life cycle of *Coenagrion puella* (Odonata : Zygoptera) from an Austrian pond. Fresh water Biology, 14 : 385 – 399.

دراسة مختبرية لدورة حياة النوع

ISCHNURA EVANSI (MORTON)(ODONATA :
COENAGRIONIDAE)

من بركة في البصرة

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الخلاصة

درست دورة حياة الرعاش الصغير *Ischnura evansi* في الظروف المختبرية وبينت النتائج ان الأناث تضع بيضها في سيقان نبات البربين المائي *Bacopa moniera* تحت سطح الماء، و ان فقس البيض يحدث في المدى الحراري بين 12 و 40 م ، و التطور الجنيني يكتمل بمدة 13 يوماً في حرارة 12 م و 7 ايام في حرارة 40 م ه . التطور اليرقي كان سريعاً نسبياً ويرتبط معنوياً بدرجة الحرارة وان مدة الطور كانت تزداد مع تقدم العمر بينما تقل مع زيادة الحرارة . بعد الفقس تمر الحشرة بثلاثة عشر طوراً حتى تصل مرحلة البلوغ . ان المدة الكلية لقضاء مرحلتي التطور الجنيني واليرقي تتراوح ما بين 197 و 278 يوماً تحت ظرفي الحرارة 40 و 25 م على التوالي .